

Intervention in Diabetic Vascular Disease by Modulation of Growth Factors

Omar Serri and Geneviève Renier

Several growth factors have been implicated in the derangements of cellular metabolism and proliferation that occur in diabetes, eg, kidney mesangial expansion, retinal neovascular formation, and acceleration of atherosclerosis in large vessels. These phenomena contribute to the development and progression of diabetic microvascular and macrovascular disease. Pharmacological interventions aimed at reducing growth factor alterations, among other actions in diabetic vasculopathy, include a multitude of classes of drugs, such as angiotensin-converting enzyme (ACE) inhibitors, calcium antagonists, lipid-lowering drugs, and somatostatin analogs. New potential interventions, ie, antisense oligonucleotide local delivery, are being applied in growth factor research and may prove beneficial in diabetic macrovascular disease.

Copyright © 1995 by W.B. Saunders Company

A MULTITUDE of complex components of diabetes mellitus (types I and II) are responsible for initiation and progression of microvascular and macrovascular lesions. The development of diabetic vasculopathy requires the simultaneous presence of a number of molecular perturbations. It is becoming increasingly clear that enhanced autocrine and paracrine growth factor production or action is one such perturbation.¹⁻³ Growth factors that are generated in response to exogenous stimuli are also elicited in response to the in vivo nonenzymatic modification of tissue components by chronically high glucose levels.^{4,5} Moreover, oxidative stress induced by hyperglycemia accelerates the formation of advanced-glycosylation end products and glycoxidation products and stimulates growth factor production.⁶ Recent findings have supported a putative role for localized imbalances of growth factor expression in the development of diabetic vasculopathy in the kidney, the retina, and large vessels. The purpose of this brief review is to consider the potential of current and future therapeutic interventions in the modulation of growth factors in diabetic vascular disease.

DIABETIC NEPHROPATHY

The early diabetic renal anomalies are mainly enlargement of the kidneys and increased glomerular filtration rate. Advanced diabetic nephropathy is characterized by thickening of the glomerular basement membrane, mesangial cell proliferation, expansion of mesangial matrix, and finally obstruction of the capillary lumen and loss of glomerular function. Among other candidates, growth factors have been suggested to play a central role in the pathogenesis of this pattern of injury.

The insulin-like growth factor (IGF) system has been the most widely studied in early diabetic disease. Infusion of IGF-I to normal human subjects results in increased circulating IGF-I and elevation of glomerular filtration rate.⁷ IGF-I administration induces kidney growth in growth hormone (GH) and IGF-I-deficient mice and rats.⁸ IGF-I is mitogenic in vitro, stimulating DNA synthesis in primary cultures of mesangial cells.⁹ IGF-I also stimulates collagen and proteoglycan synthesis.¹⁰ IGF-I accumulates in the kidney and reaches its maximum level 24 to 48 hours after induction of experimental diabetes in the rat, and then returns to normal within 4 days.¹¹ Both renal hypertrophy and IGF-I accumulation are prevented by strict glycemic control with insulin.¹¹ Accumulation of IGF-I in the kidney

is not due to increased local production, but is probably the result of increased uptake of circulating IGF-I by renal IGF-binding proteins (IGF-binding protein-1).¹² However, if IGF-I is relevant to the initiation of early diabetic nephropathy, it cannot account for the progression toward advanced nephropathy. Indeed, transgenic mice overexpressing IGF-I develop glomerular hypertrophy but not glomerulosclerosis,¹³ suggesting that other factors may be involved in the progression toward glomerulosclerosis.

The dominant histological feature of diabetic nephropathy is expansion of the extracellular matrix in the mesangium of the glomeruli, with resulting glomerulosclerosis and obliteration of the capillary surface area for filtration.¹⁴ Glomerulosclerosis can be induced by in vivo transfection of transforming growth factor- β (TGF- β) into the rat kidney.¹⁵ In vitro, TGF- β is expressed by glomerular endothelial, epithelial, and mesangial cells.¹⁶ Exposure to high glucose concentrations doubles TGF- β mRNA in cultured mesangial cells.¹⁷ Nakamura et al¹⁸ and Yamamoto et al¹⁹ have recently reported that in glomeruli of streptozotocin-diabetic rats, there is a progressive increase in the expression of TGF- β mRNA and TGF- β protein. Matrix proteins induced by TGF- β such as fibronectin, tenascin, and the proteoglycan, biglycan, were also increased.¹⁹ In glomeruli from humans with advanced diabetic nephropathy, TGF- β and fibronectin were also found to be increased, suggesting that increased TGF- β expression in both experimental and human diabetes contributes to matrix accumulation.¹⁹

Recently, numerous studies have reported the growth-modulating properties of angiotensin II (Ang II). It is now recognized that the systemic vasculature contains and can synthesize all components of the renin-angiotensin system.^{20,21} Ang II is mitogenic for a variety of cell types, including fibroblasts, vascular smooth muscle cells (SMC), and renal tubular epithelium.^{22,23} In vascular SMC, Ang II increases *c-myc* and *c-fos* transcripts, as well as mRNAs for platelet-derived growth factor (PDGF) and TGF- β .^{24,25} Infusion of Ang II into the renal artery of rats increases

From the Metabolic Unit, Research Center, Notre-Dame Hospital, Montreal, Quebec, Canada.

Address reprint requests to Omar Serri, MD, PhD, Metabolic Unit, Pavilion Mailloux, 9th Floor, Notre-Dame Hospital, 1560, Sherbrooke St East, Montreal, Quebec H2L 4M1.

Copyright © 1995 by W.B. Saunders Company

0026-0495/95/4410-4014\$03.00/0

proto-oncogene levels in whole renal cortex.²⁶ Ang II also promotes extracellular matrix accumulation and mesangial cell hypertrophy or hyperplasia.²⁷

Angiotensin-Converting Enzyme Inhibitors

Angiotensin-converting enzyme (ACE) inhibitors inhibit glomerular growth in the maturing rat kidney, an action that is independent of systemic hemodynamic changes.²⁸ In animals with experimental diabetes induced by streptozotocin, ACE inhibitors reverse the glomerular capillary hypertension and minimize the mild sclerosis that characterizes this model.²⁹ Long-term ACE-inhibition therapy is efficient in postponing the development of diabetic nephropathy in normotensive type I diabetic patients with persistent microalbuminuria.³⁰ The effect of ACE inhibition in the diabetic kidney on growth factor expression is not well known. In the remnant-kidney model, Ang II receptor antagonist prevented the development of early glomerulosclerosis and decreased PDGF expression.³¹

Calcium Antagonists

Calcium channel blockers enhance glomerular filtration rate by antagonizing the effect of vasoactive substances at the level of the afferent renal arteriole.^{32,33} Additional renal effects include attenuation of the mitogenic action of PDGF and of renal hypertrophy and reduction of mesangial entrapment of macromolecules induced by Ang II.^{34,35} However, the potential effect of calcium antagonists on growth factors in the diabetic kidney and in the long-term treatment of diabetic patients with incipient nephropathy awaits further studies.

Somatostatin Analogs

Somatostatin and its analogs have a well-known antiproliferative effect that may be due to antagonism of the actions of growth factors. The effect is mediated through a tyrosine phosphatase receptor, which in turn acts by dephosphorylating the second-messenger products of the tyrosine kinase family of some growth factor receptors.³⁶ Furthermore, somatostatin analogs can inhibit synthesis of some growth factors. Flyvbjerg et al³⁷ have shown that in diabetic rats, octreotide, a somatostatin analog, reduces serum and kidney IGF-I levels and prevents kidney hypertrophy without altering glycemic control. Serri et al³⁸ have shown that in type I diabetic patients, long-term octreotide treatment reduces kidney hyperfiltration and hypertrophy. The effect was associated with a pronounced reduction in circulating IGF-I levels, independently of GH suppression. The suggestion that octreotide may directly inhibit IGF-I synthesis at the hepatic level has later been confirmed.³⁹

Antagonists of TGF- β

Although there has been no study on the modulation of TGF- β expression in experimental or human diabetic nephropathy, there is indirect evidence that targeting TGF- β expression or action reduces matrix accumulation, thus preventing glomerulosclerosis. In a rat model of experimental glomerulonephritis, administration of anti-TGF- β reduces matrix accumulation in glomeruli.⁴⁰ Re-

cently, Border et al⁴¹ treated similar nephritic rats with repeated injections of decorin, a normal component of extracellular matrix. Decorin treatment inhibited TGF- β_1 , TGF- β_2 , and TGF- β_3 and resulted in prevention of accumulation of extracellular matrix in the glomeruli.⁴¹ The finding that dietary protein restriction in a rat model of nondiabetic glomerulosclerosis reduces both extracellular matrix expansion and glomerular expression of TGF- β ⁴² may also have important therapeutic implications in this respect. However, there may be limitations in the use of pharmacological antagonists of TGF- β because of the pleiotropic actions of this growth factor. To be therapeutically useful, the antagonist should recognize mesangium with a selectivity sufficiently greater than that with which it recognizes normal sources of TGF- β .

DIABETIC RETINOPATHY

The principal hypothesis concerning the molecular basis of proliferative diabetic retinopathy has focused on the presence of diffusible angiogenic growth factors that are released from the retina under hyperglycemic conditions. The formation of new blood vessels involves at least (1) degradation of the extracellular matrix surrounding normal capillaries, (2) endothelial migration and proliferation, (3) endothelial tube formation, and (4) anastomosis of nascent tubes. However, the nature of growth-promoting mediators implicated in these phenomena is not well known. Numerous factors have been identified that can elicit new blood vessel formation in angiogenesis assays. The only data available from human studies have been limited to measurements of growth factors in intraocular fluid samples from patients with diabetic proliferative retinopathy. Attempts to correlate circulating IGF-I levels and proliferative diabetic retinopathy have yielded conflicting results.⁴³⁻⁴⁷ Vitreous concentrations of IGF-I may be of more importance than systemic levels. Grant et al⁴⁸ found markedly elevated IGF-I levels in the vitreous from patients with proliferative diabetic retinopathy undergoing vitrectomy as compared with nondiabetic patients.

Basic fibroblast growth factor (bFGF) is one potentially important growth factor that can initiate mitogenesis in endothelial cells. It is stored in high concentrations within the extracellular matrix as an inactive complex, and is released when the matrix is dissolved by activated endothelial cells. Sivalingam et al⁴⁹ reported elevated levels of bFGF in vitrectomy samples from patients with active proliferative diabetic retinopathy.

Microvascular endothelial cells also synthesize and respond to PDGF. PDGF, which stimulates endothelial migration but not proliferation, is able to elicit a full angiogenic response in the rabbit corneal assay.⁵⁰

In vitro, IGF-I and bFGF synergistically stimulate growth of human retinal endothelial cells.⁵¹

Somatostatin Analogs

Octreotide inhibits the in vitro growth of human retinal endothelial cells induced by IGF-I and bFGF.⁵¹ Furthermore, somatostatin analogs have been shown to inhibit angiogenesis in the chick allantoic membrane.⁵²

In a controlled trial, a 1-year continuous subcutaneous infusion of octreotide had no significant effect on early retinopathy in type I diabetic patients.⁵³ In four patients with type I diabetes and severe proliferative retinopathy, long-term treatment (6 to 20 months) with octreotide led to stabilization of proliferative retinopathy.⁵⁴ In one controlled study,⁵⁵ two of eight patients demonstrated improvement of proliferative retinopathy with BIM23014, a somatostatin analog, whereas control patients showed worsening of fluorescein leakage. The other six patients receiving somatostatin analog infusion showed stabilization of lesions.⁵⁵ In these studies, circulating IGF-I levels were inhibited by somatostatin analog treatment. Randomized trials are still lacking to conclude on the efficacy of these analogs. However, it appears that they cannot replace photocoagulation therapy, but rather constitute in selected cases an adjunct therapy to photocoagulation, at least in advanced proliferative lesions. The apparent limited efficacy of these drugs may be due to their use in advanced stages of retinopathy, in which damage may be irreversible. In considering future trials, timing and duration of treatment should aim to prevent or at least slow angiogenesis at preproliferative stages of retinopathy.

ACE Inhibitors

Treatment with ACE inhibitors has been shown to reduce fluorescein leakage in diabetic subjects with background retinopathy.⁵⁶ In a small series of patients, a 2-year double-blinded trial also showed decreased retinal deterioration and some improvement in normotensive diabetic subjects treated with ACE inhibitors.⁵⁷

Miscellaneous

Thalidomide is a well-known potent teratogen. It was postulated that its teratogenicity may be related to an inhibition of blood vessel growth in the developing fetal limb bud.⁵⁸ The effect of thalidomide was examined on growing vasculature in the chicken chorioallantoic membrane and in the rabbit cornea.⁵⁸ The study showed that thalidomide is an inhibitor of angiogenesis induced by bFGF.

A novel endogenous angiogenesis inhibitor has been recently reported.⁵⁹ Angiostatin, which is a 38-kd fragment of plasminogen, potently inhibits *in vitro* endothelial proliferation and blocks bFGF-induced neovascularization in the chick chorioallantoic membrane. Future studies should determine whether thalidomide and angiostatin can be useful in the treatment of patients with proliferative diabetic retinopathy.

ATHEROSCLEROSIS

Atherosclerosis is a response of the artery wall to a variety of initiating agents with multiple pathogenic mechanisms contributing to the formation of the atherosclerotic plaque. One major transitional event in lesion progression is the migration to and proliferation within the intima of medial SMC, leading to the synthesis of plaque collagens, elastin, and proteoglycans that contribute further to lesion progression. Molecules controlling smooth muscle growth

are derived from platelets, monocyte/macrophages, T-lymphocytes, endothelial cells, and SMC themselves. They include PDGF, TGF- β , bFGF, heparin-binding epidermal growth factor (EGF), IGF-I, interleukin-1 (IL-1), and tumor necrosis factor alpha (TNF α).

Examination of human atherosclerotic specimens has revealed mRNAs transcribed from genes for a variety of growth-promoting substances not normally expressed in arterial wall. A marked increase of PDGF has been demonstrated in SMC and adjacent macrophages in developing atherosclerotic plaques. SMC are the predominant source of PDGF-A chain,⁶⁰ whereas macrophages express mRNA for PDGF-B chain.⁶¹ Even though PDGF is neither a necessary nor sufficient contributor to human atherogenesis, it is the prototypic SMC mitogen. PDGF by itself is a powerful chemoattractant, even in submitogenic concentrations, and is able to induce SMC migration more rapidly than proliferation.⁶² It induces extensive SMC replication in conjunction with other serum factors.

IGF-I mRNA levels are also markedly increased in plaques, and *in situ* hybridization studies have shown that IGF-I mRNA is expressed by SMC themselves. An increase in vascular load is associated with an increase in IGF-I production by endothelial cells and SMC.⁶³ Moreover, enhanced IGF-I expression has also been observed during vessel restenosis and angiogenesis.⁶⁴ Because PDGF and IGF-I may function together to result in enhanced rates of SMC proliferation *in vitro*, localization of their respective transcripts in vessel-wall cell types suggests that they also have the potential to stimulate lesion development.

Recent studies have supported the involvement of FGFs in vascular pathobiology. These growth factors can foster re-endothelialization,⁶⁵ mediate SMC replication in balloon-injured arteries,⁶⁶ and possess chemotactic activity.⁶⁷ Increased levels of acidic FGF mRNA have been demonstrated in atherosclerotic plaques, with most of the hybridization signal for this growth factor being localized in macrophages.⁶⁸

Although detected in minor amounts in normal arterial wall, TGF- β mRNA expression is markedly increased during atherosclerosis.⁶⁹ TGF- β exerts a powerful control over SMC proliferation.^{70,71} During the early stages of SMC proliferation, TGF- β appears to be antiproliferative,⁷⁰ whereas later in the development of the fibrous plaque, TGF- β is growth-stimulatory to SMC.⁷² It also strongly stimulates SMC to produce the components of the extracellular matrix.⁷³ By promoting angiogenesis, it may also account for the plaque disruption of late atherosclerosis.⁷⁴

Activated monocytes release large amounts of several cytokines with growth-promoting properties in the arterial wall, including IL-1 and TNF α . Although IL-1 *per se* is not mitogenic for SMC, it induces autocrine expression and release of PDGF-AA from SMC.⁷² IL-1 may also facilitate SMC migration⁷² and induce SMC to synthesize some components of the extracellular matrix.⁷⁵ TNF α is one of the earliest-acting atherosclerotic cytokines. It stimulates SMC to release IL-1⁷⁶ and facilitates recruitment of monocytes in the arterial wall. It is also responsible for the later development of plaque neovascularization.⁷⁷

Several key components of atherosclerotic plaque initiation are likely to be enhanced by diabetes, including intimal lipoprotein influx and accumulation, monocyte-macrophage recruitment, generation of free radicals, and lipoprotein oxidation.⁷⁸ Diabetes can also probably contribute to plaque progression by augmented SMC proliferation or connective tissue synthesis. In this regard, advanced-glycosylation end products may play a central role by enhancing PDGF and IGF-I secretion in the vascular wall.⁴

Pharmacological Modulation of Growth Factors in Atherosclerosis

Decreasing elevated total and low-density lipoprotein (LDL) cholesterol levels is a well-established therapeutic method of manipulating plasma lipids to reduce the progression of atherosclerosis. Lipid-lowering drugs are numerous and include nicotinic acid, bile acid sequestrants, fibric acid derivatives, hydroxymethyl glutaryl coenzyme A (HMG CoA) reductase inhibitors, and probucol. In addition to their effects on lipid metabolism, some of these drugs also possess antiproliferative properties.

Fibric Acid Derivatives

A role of fenofibric acid as a PDGF antagonist was proposed in 1983 by Pascal et al,⁷⁹ who reported an inhibitory effect of this drug on DNA synthesis induced by a PDGF-rich platelet extract in cultured rat vascular SMC. Similar inhibitory effects of fenofibrate but not of fenofibric acid on human vascular SMC growth have been recently documented by Munro et al.⁸⁰ The antiproliferative effect of fibric acid derivatives could account for the regression of coronary atherosclerotic plaques observed following fenofibrate treatment.⁸¹

HMG CoA Reductase Inhibitors

Inhibition of HMG CoA reductase, the enzyme responsible for biosynthesis of mevalonate, can result in cell cycle arrest.⁸² It has been reported that lovastatin, one inhibitor of HMG CoA reductase, may increase the frequency of regression and reduce the progression of coronary atherosclerosis by combination therapy with colestipol in clinical studies.⁸³ This drug also has been shown to reduce intimal hyperplasia after balloon angioplasty in the hypercholesterolemic atherosclerotic rabbit.⁸⁴ It has been proposed that lovastatin exerts these effects by inhibiting SMC proliferation. Lovastatin also attenuates induction of *c-fos* mRNA by EGF, insulin, and IGF-I, but not by bFGF or PDGF.⁸⁵ In a similar way, mevastatin also suppresses *c-fos* and *c-myc* mRNA accumulation in response to serum stimulation,⁸⁶ demonstrating that a component of the early response to growth factors is sensitive to mevalonate deprivation. Finally, inhibition of cultured vascular SMC migration by simvastatin, another HMG CoA inhibitor, has also been reported.⁸⁷ Although the mechanisms by which these drugs exert their effects are unclear, it appears that HMG CoA reductase inhibitors can attenuate activation of the cells within the vessel wall. Further studies are required to elucidate the importance of this property in the prevention of atherosclerotic lesions.

Calcium Antagonists

Experimental and clinical evidence suggests that calcium antagonists may be able to prevent or retard the progression of atherosclerosis by mechanisms independent of and in addition to blood pressure reduction.⁸⁸ A wide range of explorations for the apparent antiatherosclerotic effects of calcium antagonists have been offered. Some of these drugs may suppress vascular SMC migration⁸⁹ and proliferation.⁹⁰ They have been reported to reduce PDGF-induced growth of SMC and to inhibit Ang II- and PDGF-BB-induced DNA synthesis in the cells.⁹¹⁻⁹³ A recent study provides evidence that the antiproliferative effect of isradipine on PDGF- and Ang II-induced vascular SMC growth may be due to inhibition of expression of the transcriptional factor, *c-fos*.⁹³ Some calcium channel blockers such as flunarizine may also enhance transcription of many genes induced by PDGF-BB such as LDL receptor, HMG CoA reductase, *c-fos*, *c-jun*, and cytokines.^{94,95}

ACE Inhibitors

Ang II may play an important role in modulation of mechanisms underlying SMC proliferation by inducing the genes encoding growth factors and stimulating the expression of factors affecting activities of these cells.^{96,97} A role of the local renin-angiotensin system in the myointimal proliferative processes that develop in the vascular wall is supported by several observations. It has been shown that Ang II is a potent growth factor for vascular SMC in vitro,⁹⁸ and that ACE inhibitors prevent or attenuate development of myointimal hyperplasia after endothelial denudation and vascular injury.^{96,99} Some ACE inhibitors blunt PDGF-BB-stimulated de novo DNA synthesis and cell proliferation in vascular SMC,¹⁰⁰ and decrease de novo synthesis of mRNA of the transcription factors *c-fos* and *c-jun* achieved by PDGF-BB.¹⁰⁰ Finally, they intensify the transcription of LDL receptor mRNA induced by PDGF-BB.¹⁰⁰ Despite these experimental evidences, the clinical importance of such findings is still controversial and needs to be assessed in future investigations.

Miscellaneous

Although antioxidants, including probucol, exert their antiatherosclerotic effect by preventing oxidation of LDL, their ability to directly inhibit growth factor biosynthesis and SMC proliferation is an additional property that may contribute to their antiatherosclerotic effects. Alpha-tocopherol has been demonstrated to inhibit growth factor-induced SMC proliferation.¹⁰¹ In addition, alpha-tocopherol and probucol depress IL-1 β expression.¹⁰²

Octreotide decreases chemotaxis of monocytes induced by GH in vitro¹⁰³ and inhibits release of superoxide anion from stimulated monocytes.¹⁰⁴ Octreotide has also recently been reported to reduce IGF-I- and bFGF-induced human coronary artery SMC proliferation.¹⁰⁵ Its potential clinical usefulness in reducing the incidence of restenosis remains to be evaluated.

Table 1 summarizes the principal data for growth factor modulation by different pharmacological interventions in diabetic vascular disease.

Table 1. Principal Data of Growth Factor Modulation by Pharmacological Interventions in Diabetic Vascular Disease

Agents	Nephropathy	Retinopathy	Atherosclerosis
ACE inhibitors	<ul style="list-style-type: none"> ● Inhibit glomerular growth in maturing rat kidney²⁸ ● Reverse glomerular capillary hypertension in experimental diabetes, and minimize glomerulosclerosis²⁹ 	<ul style="list-style-type: none"> ● Reduce fluorescein leakage in background retinopathy⁵⁶ ● Decrease retinal deterioration in normotensive diabetic subjects⁵⁷ 	<ul style="list-style-type: none"> ● Prevent or attenuate myointimal hyperplasia^{96,99} ● Can blunt PDGF-stimulated SMC proliferation¹⁰⁰
Calcium antagonists	<ul style="list-style-type: none"> ● Antagonize the effect of vasoactive substances at afferent renal arteriole level^{32,33} ● Attenuate the mitogenic action of PDGF and renal hypertrophy, and attenuate mesangial entrapment of macromolecules induced by Ang II^{34,35} 		<ul style="list-style-type: none"> ● Can suppress SMC migration and proliferation^{89,90} ● Reduce stimulated SMC proliferation by PDGF and inhibit stimulated DNA synthesis by Ang II in SMC⁹¹⁻⁹³
Somatostatin analogus	<ul style="list-style-type: none"> ● Reduce kidney hyperfiltration³⁸ ● Reduce kidney hypertrophy^{37,38} ● Inhibit circulating IGF-I levels^{37,38} ● Reduce kidney IGF-I accumulation³⁷ 	<ul style="list-style-type: none"> ● Inhibit growth of human retinal endothelial cells induced by IGF-I and bFGF⁵¹ ● Inhibit angiogenesis in chick allantoic membrane⁵² ● Improve or stabilize proliferative retinopathy in small series of patients^{54,55} ● Inhibit circulating IGF-I levels^{54,55} 	<ul style="list-style-type: none"> ● Decrease chemotaxis of monocytes induced by GH in vitro¹⁰³ ● Inhibit release of superoxide anion from stimulated monocytes¹⁰⁴ ● Inhibit human coronary artery SMC proliferation induced by IGF-I and bFGF¹⁰⁵
HMG CoA reductase inhibitors			<ul style="list-style-type: none"> ● Have the potential to inhibit SMC proliferation^{84,87} ● Lovastatin and simvastatin Arrest cells in the G1 phase⁸² Attenuate induction of <i>c-fos</i> by EGF, insulin, and IGF-I⁸⁵
Potential future interventions	<ul style="list-style-type: none"> ● Antagonists of growth factors Anti-TGF-β⁴⁰ Decorin⁴¹ ● Antagonists of Ang II receptors³¹ 	<ul style="list-style-type: none"> ● Inhibitors of angiogenesis Thalidomide⁵⁸ Angiostatin⁵⁹ 	<ul style="list-style-type: none"> ● Antagonists of growth factors ● Antisense oligonucleotides to proto-oncogenes¹⁰⁶

FUTURE PROSPECTS FOR GROWTH FACTOR-MODULATING THERAPEUTIC STRATEGIES

It is still unclear which growth factor may be involved as a primary initiator of each specific diabetic vasculopathy and which one is a secondary but still important mediator of injury. It seems unlikely that blocking one mediator alone will reverse growth perturbations when other pathways remain active. Nevertheless, the possibility of treatment using anti-growth factor antibodies or growth factor receptor blockers should be considered. The former approach is unlikely to succeed because the dilution of the antibody will not be able to significantly reduce local production of a growth factor. Receptor blockade is more specific and more promising. Among other therapeutic interventions, a classic pharmacological approach can evaluate combinations of different classes of drugs such as ACE inhibitors or calcium

antagonists with somatostatin analogs (when oral preparations are available) to confer additional benefits in the management of progressive diabetic nephropathy. Inhibition of angiogenesis in diabetic proliferative retinopathy can also be evaluated with new drugs such as thalidomide or angiostatin. A future therapeutic strategy targeting the vasculature could be local delivery of antisense oligonucleotides to inhibit gene products implicated in SMC growth.¹⁰⁶ This may prove beneficial in the treatment of acute atherosclerotic processes in the diabetic patient, such as restenosis following coronary artery angioplasty or bypass grafts.

ACKNOWLEDGMENT

The authors thank Dr Eugenio Rasio for constructive comments, and Fernanda Janeiro for secretarial assistance in preparation of the manuscript.

REFERENCES

1. Clemmons DR: Role of peptide growth factors in development of macrovascular complications of diabetes. *Diabetes Care* 14:153-156, 1991
2. Schwieger J, Fine LG: Renal hypertrophy, growth factors, and nephropathy in diabetes mellitus. *Semin Nephrol* 10:242-253, 1990
3. Woolf AS, Bosch RJ, Fine LG: Growth factors in the pathogenesis of renovascular complications of diabetes mellitus. *J Hypertens* 10:S11-S16, 1992
4. Kirstein M, Brett J, Radoff S, et al: Advanced protein glycosylation induces transendothelial human monocyte chemotaxis and secretion of platelet-derived growth factor: Role in vascular disease of diabetes and aging. *Proc Natl Acad Sci USA* 87:9010-9014, 1990

5. Doi T, Vlassara H, Kirstein M, et al: Receptor-specific increase in extracellular matrix production in mouse mesangial cells by advanced glycosylation end products is mediated via platelet-derived growth factor. *Proc Natl Acad Sci USA* 89:2873-2877, 1992
6. Baynes JW: Role of oxidative stress in development of complications in diabetes. *Diabetes* 40:405-412, 1991
7. Guler HP, Schmid C, Zapf J, et al: Effects of recombinant insulin-like growth factor I on insulin secretion and renal function in normal human subjects. *Proc Natl Acad Sci USA* 86:2868-2872, 1989
8. Hirschberg R, Kopple JD: Evidence that IGF-I increases renal plasma flow and glomerular filtration rate in fasted rats. *J Clin Invest* 83:326-330, 1989
9. Lowe WL Jr: Biological actions of the insulin-like growth factors, in LeRoith D (ed): *Insulin-Like Growth Factors: Molecular and Cellular Aspects*. Boca Raton, FL, CRC, 1991, pp 49-85
10. Bar RS, Dake BL, Stueck S: Stimulation of proteoglycans by IGF-I and II in microvessel and large vessel endothelial cells. *Am J Physiol* 253:E21-E27, 1987
11. Flyvbjerg A, Bornfeldt KE, Marshall SM, et al: Kidney IGF-I mRNA in initial renal hypertrophy in experimental diabetes in rats. *Diabetologia* 33:334-338, 1990
12. LeRoith D, Werner H, Phillip M, et al: The role of insulin-like growth factors in diabetic kidney disease. *Am J Kidney Dis* 22:722-726, 1993
13. Doi T, Striker LJ, Quaife C, et al: Progressive glomerulosclerosis develops in transgenic mice chronically expressing growth hormone and growth hormone releasing factor but not in those expressing insulin-like growth factor-1. *Am J Pathol* 131:398-403, 1988
14. Mauer SM, Steffes MW, Ellis EN, et al: Structural-functional relationships in diabetic nephropathy. *J Clin Invest* 74:1143-1155, 1984
15. Isaka Y, Fujiwara Y, Ueda N, et al: Glomerulosclerosis induced by in vivo transfection of transforming growth factor- β or platelet-derived growth factor gene into the rat kidney. *J Clin Invest* 92:2597-2601, 1993
16. Mackay K, Kondaiah P, Dalielpour D, et al: Expression of transforming growth factor- β 1 and β 2 in rat glomeruli. *Kidney Int* 38:1095-1100, 1990
17. Kaname S, Uchida S, Ogato E, et al: Autocrine secretion of transforming growth factor- β in cultured rat mesangial cells. *Kidney Int* 42:1319-1327, 1992
18. Nakamura T, Fukui M, Ebihara I, et al: mRNA expression of growth factors in glomeruli from diabetic rats. *Diabetes* 42:450-456, 1993
19. Yamamoto T, Nakamura T, Noble NA, et al: Expression of transforming growth factor β is elevated in human and experimental diabetic nephropathy. *Proc Natl Acad Sci USA* 90:1814-1818, 1993
20. Lynch DR, Peach MJ: Molecular biology of angiotensinogen. *Hypertension* 17:263-269, 1991
21. Owens GK: Control of hypertrophic versus hyperplastic growth of vascular smooth muscle cells. *Am J Physiol* 257:H1755-H1765, 1989
22. Schilling P, Fischer H, Ganten D: Angiotensin and cell growth: A link to cardiovascular hypertrophy. *J Hypertens* 9:3-15, 1991
23. Wolf G, Neilson EG: Angiotensin II induces cellular hypertrophy in cultured murine proximal tubular cells. *Am J Physiol* 259:F768-F777, 1990
24. Naftilan AJ, Pratt RE, Dzau VJ: Induction of platelet derived growth factor A-chain and *c-myc* gene expressions by angiotensin II in cultured rat vascular smooth muscle cells. *J Clin Invest* 83:1419-1424, 1989
25. Powell S, Rouge M, Muler RKM, et al: Cilazapril suppresses myointimal proliferation after vascular injury: Effects of growth factor induction on vascular smooth muscle. *Basic Res Cardiol* 86:65-74, 1991 (suppl 1)
26. Rosenberg ME, Hostetter TH: Effect of angiotensin II and norepinephrine on early growth response genes in the rat kidney. *Kidney Int* 43:601-609, 1993
27. Harris R, Ichikawa I: Angiotensin actions in the kidney: Renewed insight into the old hormone. *Kidney Int* 40:583-596, 1991
28. Fogo A, Yoshida Y, Yared A, et al: Importance of angiogenic action of angiotensin II in the glomerular growth of maturing kidneys. *Kidney Int* 38:1068-1074, 1990
29. Anderson S, Rennke HG, Garcia DL, et al: Short and long-term effects of antihypertensive therapy in the diabetic rat. *Kidney Int* 36:526-536, 1989
30. Mathiesen ER, Hommel E, Giese J, et al: Efficacy of captopril in postponing nephropathy in normotensive insulin dependent diabetic patients with microalbuminuria. *Br Med J* 303:81-87, 1991
31. Tanaka R, Fogo A: Internephron heterogeneity of growth factors and sclerosis—Modulation of platelet derived growth factor (PDGF) by angiotensin II (AII). *J Am Soc Nephrol* 4:637, 1993 (abstr)
32. Epstein M, Loutzenhiser R: Effects of calcium antagonists on renal hemodynamics. *Am J Kidney Dis* 16:10-14, 1990 (suppl 1)
33. Takenaka T, Epstein M, Forster H, et al: Attenuation of endothelin effects by a chloride channel inhibitor indanyloxyacetic acid. *Am J Physiol* 262:F799-F806, 1992
34. Sweeney C, Shultz P, Raj L: Interactions of the endothelium and mesangium in glomerular injury. *J Am Soc Nephrol* 1:S13-S20, 1990
35. Raj L, Keane W: Glomerular mesangium: Its function and relationship to angiotensin II. *Am J Med* 79:24-30, 1985 (suppl 36)
36. Lee MT, Liebow C, Kamer AR, et al: Effect of epidermal growth factor and analogues of luteinizing hormone-releasing hormone and somatostatin on phosphorylation and dephosphorylation of tyrosine residues of specific protein substrates in various tumors. *Proc Natl Acad Sci USA* 88:1656-1660, 1991
37. Flyvbjerg A, Frystyk J, Thorlacius-Ussing O, et al: Somatostatin analogue administration prevents increase in kidney somatomedin C and initial renal growth in diabetic and uninephrectomized rats. *Diabetologia* 32:261-265, 1989
38. Serri O, Beauregard H, Brazeau P, et al: Somatostatin analogue, octreotide, reduces increased glomerular filtration rate and kidney size in insulin-dependent diabetes. *JAMA* 265:888-892, 1991
39. Serri O, Brazeau P, Kachra Z, et al: Octreotide inhibits insulin-like growth factor-I hepatic gene expression in the hypophysectomized rat: Evidence for a direct and indirect mechanism of action. *Endocrinology* 130:1816-1821, 1992
40. Border WA, Okuda S, Languino L, et al: Suppression of experimental glomerulonephritis by antiserum against transforming growth factor β 1. *Nature* 346:371-374, 1990
41. Border WA, Noble NA, Yamamoto T, et al: Natural inhibitor of transforming growth factor- β protects against scarring in experimental kidney disease. *Nature* 360:361-364, 1992
42. Okuda S, Nakamura T, Yamamoto T, et al: Dietary protein restriction rapidly reduces transforming growth factor β -1 expression in experimental glomerulonephritis. *Proc Natl Acad Sci USA* 88:9765-9769, 1991
43. Merimee TJ, Zapf J, Froesch ER: Insulin-like growth

factors—Studies in diabetics with and without retinopathy. *N Engl J Med* 309:527-530, 1983

44. Lambertson RP, Goodman AD, Kassoff A, et al: Von Willebrand factor (VIII:Ag), fibronectin, and insulin-like growth factors I and II in diabetic retinopathy and nephropathy. *Diabetes* 33:125-129, 1984

45. Hyer SL, Sharp PS, Brooks RA, et al: Serum IGF-I concentration in diabetic retinopathy. *Diabetic Med* 5:356-360, 1988

46. Nardelli GM, Guastamacchia E, DiPaolo S, et al: Somatomedin-C (Sm-C): Study in diabetic patients with and without retinopathy. *Acta Diabetol Lat* 26:217-224, 1989

47. Dills DG, Moss SE, Klein R, et al: Association of elevated IGF-I levels with increased retinopathy in late-onset diabetes. *Diabetes* 40:1725-1730, 1991

48. Grant M, Russel B, Fitzgerald C, et al: Insulin-like growth factors in vitreous. Studies in control and diabetic subjects with neovascularization. *Diabetes* 35:416-420, 1986

49. Sivalingam A, Kenney J, Brown GC, et al: Basic fibroblast growth factor levels in the vitreous of patients with proliferative diabetic retinopathy. *Arch Ophthalmol* 108:869-872, 1990

50. Beitz JG, Kim I-S, Calabresi P, et al: Human microvascular endothelial cells express receptors for platelet derived growth factor. *Proc Natl Acad Sci USA* 88:2021-2025, 1991

51. Grant MB, Caballero S, Millard WJ: Inhibition of IGF-I and b-FGF stimulated growth of human retinal endothelial cells by the somatostatin analogue, octreotide: A potential treatment for ocular neovascularization. *Regul Pept* 48:267-278, 1993

52. Woltering EA, Barrie R, O'Doriso TM, et al: Somatostatin analogues inhibit angiogenesis in the chick allantoic membrane. *J Surg Res* 50:245-251, 1991

53. Kirkegaard C, Norgaard K, Sndrgaard O, et al: Effect of one year continuous subcutaneous infusions of a somatostatin analogue, octreotide, on early retinopathy, metabolic control and thyroid function in type I (insulin-dependent) diabetes mellitus. *Acta Endocrinol (Copenh)* 122:766-772, 1990

54. Mallet B, Vialettes B, Haroche S, et al: Stabilization of severe proliferative diabetic retinopathy by long-term treatment with SMS 201-995. *Diabetes Metab* 18:438-444, 1992

55. McCombe M, Lightman S, Eckland DJ, et al: Effect of a long-acting somatostatin analogue (BIM23014) on proliferative diabetic retinopathy: A pilot study. *Eye* 5:569-575, 1991

56. Jackson WE, Holmes DL, Garg SK, et al: Angiotensin-converting enzyme inhibitor therapy and diabetic retinopathy. *Ann Ophthalmol* 24:99-103, 1992

57. Chase HP, Garg SK, Harris S, et al: Angiotensin-converting enzyme inhibitor treatment for young normotensive diabetic subjects: A two-year trial. *Ann Ophthalmol* 25:284-289, 1993

58. D'Amato RJ, Loughnan MS, Flynn E, et al: Thalidomide is an inhibitor of angiogenesis. *Proc Natl Acad Sci USA* 91:4082-4085, 1994

59. O'Reilly MS, Holmgren L, Shing Y, et al: Angiostatin: A novel angiogenesis inhibitor that mediates the suppression of metastases by a Lewis lung carcinoma. *Cell* 79:315-328, 1994

60. Wilcox JN, Smith KM, Williams LT, et al: Platelet-derived growth factor mRNA detection in human atherosclerotic plaques by in situ hybridization. *J Clin Invest* 82:1134-1143, 1988

61. Ross R, Masuda J, Raines EW, et al: Localization of PDGF- β protein in macrophages in all phases of atherogenesis. *Science* 248:1009-1012, 1990

62. Grotendorst GR, Chang T, Seppa HEJ, et al: Platelet-derived growth factor is a chemoattractant for vascular smooth muscle cells. *J Cell Physiol* 113:261-266, 1982

63. Hansson HA, Jennische E, Skottnier A: IGF-I expression in blood vessels varies with vascular load. *Acta Physiol Scand* 129:165-169, 1987

64. Hansson HA, Brandsten C, Lossing C, et al: Transient expression of insulin-like growth factor 1 immunoactivity by vascular cells during angiogenesis. *Exp Mol Pathol* 50:125-138, 1989

65. Lindner V, Majack RA, Reidy MA: Basic fibroblast growth factor stimulated endothelial regrowth and proliferation in denuded arteries. *J Clin Invest* 85:2004-2008, 1990

66. Lindner V, Lappi DA, Baird A, et al: Role of basic fibroblast growth factor in vascular lesion formation. *Circ Res* 68:106-113, 1991

67. Sato Y, Rifkin DB: Autocrine activities of basic fibroblast growth factor: Regulation of endothelial cell movement, plasminogen activator synthesis and DNA synthesis. *J Cell Biol* 107:1199-1205, 1988

68. Brogi E, Winkles JA, Underwood R, et al: Distinct patterns of expression of fibroblast growth factors and their receptors in human atheroma and non atherosclerotic arteries. *J Clin Invest* 92:2408-2418, 1993

69. Nikol S, Isner J, Pickering G, et al: Transforming growth factor- β 1: A peptide growth factor with increased expression in vascular restenosis. *J Am Coll Cardiol* 19:323A, 1992 (abstr)

70. Morisaki U, Kowano M, Koyama N, et al: Effects of transforming growth factor- β 1 on growth of aortic smooth muscle cells. *Atherosclerosis* 88:227-234, 1991

71. Schlumberger W, Thie M, Rauterberg J, et al: Collagen synthesis in cultured aortic smooth muscle cells: Modulation by collagen lattice culture, transforming growth factor- β 1 and epidermal growth factor. *Arterioscler Thromb* 11:1660-1666, 1991

72. Raines EW, Dower SK, Ross R: Interleukin-1 mitogenic activity for fibroblasts and smooth muscle cells is due to PDGF-AA. *Science* 243:393-396, 1989

73. Olson EN, Sternberg E, Hu JS, et al: Regulation of myogenic differentiation by type β transforming growth factor. *J Cell Biol* 103:1799-1805, 1986

74. Roberts AB, Sporn MB, Assoian RK, et al: Transforming growth factor type β : Rapid induction of fibrosis and angiogenesis in vivo and stimulation of collagen formation in vitro. *Proc Natl Acad Sci USA* 83:4167-4171, 1986

75. Amento EP, Ehsani N, Palmer H, et al: Cytokines and growth factors positively and negatively regulate interstitial collagen gene expression in human vascular smooth muscle. *Arterioscler Thromb* 11:1223-1230, 1991

76. Warner SJC, Libby P: Human vascular smooth muscle cells: Target for and source of tumour necrosis factor. *J Immunol* 142:100-109, 1990

77. Barath P, Fishbein MC, Cao J, et al: Detection and localization of tumour necrosis factor in human atheroma. *Am J Cardiol* 65:297-302, 1990

78. Schwartz CJ, Valente AJ, Sprague EA, et al: Pathogenesis of the atherosclerotic lesion. Implications for diabetes mellitus. *Diabetes Care* 15:1156-1167, 1992

79. Pascal M, Sepulchre C, Chazan JB, et al: Evidence for the inhibition of platelet-derived growth factor induced rat smooth muscle cells DNA synthesis by fenofibrate acid at the Go/G1 cell cycle level. *Life Sci* 33:925-933, 1983

80. Munro E, Patel M, Chan P, et al: Growth inhibition of human vascular smooth muscle cells by fenofibrate: A possible therapy for restenosis. *Cardiovasc Res* 28:615-620, 1994

81. Hahmann HW, Bunte H, Hellwig N, et al: Progression and regression of minor coronary arterial narrowings by quantitative angiography after fenofibrate therapy. *Am J Cardiol* 67:957-961, 1991

82. Quesney-Huuneeus V, Wiley MH, Siperstein MD: Essential role for mevalonate synthesis in DNA replication. *Proc Natl Acad Sci USA* 76:5056-5060, 1979

83. Brown G, Albers JJ, Fisher LD, et al: Regression of

- coronary artery disease as a result of intensive lipid-lowering therapy in men with high levels of apolipoprotein B. *N Engl J Med* 323:1289-1298, 1990
84. Gellman J, Ezekowitz MD, Sarembock IJ, et al: Effect of lovastatin on intimal hyperplasia after balloon angioplasty: A study in an atherosclerotic hypercholesterolemic rabbit. *J Am Coll Cardiol* 17:251-259, 1991
 85. Vincent TS, Wulfert E, Merler E: Inhibition of growth factor signalling pathways by lovastatin. *Biochem Biophys Res Commun* 180:1284-1289, 1991
 86. Barbu V, Dautry F: Mevalonate deprivation alters the induction of *fos* and *myc* by growth factors. *Oncogene* 5:1077-1080, 1990
 87. Hidaka Y, Eda T, Yonemoto M, et al: Inhibition of cultured vascular smooth muscle cell migration by simvastatin (MK-733). *Atherosclerosis* 95:87-94, 1992
 88. Holzgreve H, Burkle B: Anti-atherosclerotic effects of calcium antagonists. *J Hypertens* 11:S55-S59, 1993 (suppl 1)
 89. Nomoto A, Hirosumi J, Sekiguchi C, et al: Antiatherosclerotic activity of FR34235 (nivaldipine), a new potent calcium antagonist. *Atherosclerosis* 64:255-261, 1987
 90. Pauletto P, Scannapieco G, Borriore AC, et al: A nifedipine-sensitive smooth muscle cell population is present in the atherosclerotic rabbit aorta. *Arterioscler Thromb* 11:928-939, 1991
 91. Nilsson J, Sjolund M, Palmerg L, et al: The calcium antagonist nifedipine inhibits arterial smooth muscle cell proliferation. *Atherosclerosis* 58:109-122, 1985
 92. Ko YD, Sachinidis A, Graack GH, et al: Inhibition of angiotensin II and platelet-derived growth factor-induced vascular smooth muscle cell proliferation by calcium entry blockers. *J Clin Invest* 70:113-117, 1992
 93. Ko Y, Totzke G, Graack GH, et al: Action of dihydropyridine calcium antagonists on early growth response gene expression and cell growth in vascular smooth muscle cells. *J Hypertens* 11:1171-1178, 1993
 94. Roth M, Keul R, Emmons LR, et al: Manidipine regulates the transcription of cytokine genes. *Proc Natl Acad Sci USA* 89:4071-4075, 1992
 95. Roth M, Emmons LR, Perruchoud AP, et al: Expressions of the low density lipoprotein receptor and 3-hydroxy-3-methylglutaryl coenzyme A reductase genes are stimulated by recombinant platelet-derived growth factor isomers. *Proc Natl Acad Sci USA* 88:1888-1892, 1991
 96. Powell JS, Muller RKM, Rouge M, et al: The proliferative response to vascular injury is suppressed by angiotensin-converting enzyme inhibition. *J Cardiovasc Pharmacol* 16:S42-S49, 1990 (suppl 4)
 97. Re R: Angiotensin and the regulation of cellular growth. Pathophysiologic implications for cardiovascular and noncardiovascular tissues. *Am J Hypertens* 4:217S-219S, 1991
 98. Campbell-Boswell M, Robertson AL: Effects of angiotensin II and vasopressin on human smooth muscle cells in vitro. *Exp Mol Pathol* 35:265-276, 1981
 99. Powell JS, Muller RKM, Rouge M, et al: Suppression of the vascular response to injury: The role of angiotensin-converting enzyme inhibitors. *J Am Coll Cardiol* 17:137B-142B, 1991
 100. Block LH, Keul R, Crabos M, et al: Transcriptional activation of low density lipoprotein receptor gene by angiotensin-converting enzyme inhibitors and Ca^{2+} -channel blockers involves protein kinase C isoforms. *Proc Natl Acad Sci USA* 90:4097-4101, 1993
 101. Boscoboinik D, Szweczyk A, Azzi A: Alpha-tocopherol (vitamin E) regulates vascular smooth muscle cell proliferation and protein kinase C. *Arch Biochem Biophys* 286:264-269, 1991
 102. Akesson AL, Woods CW, Mosher LB, et al: Inhibition of IL-1 β expression in THP-1 cells by probucol and tocopherol. *Atherosclerosis* 86:261-270, 1991
 103. Wiedermann CJ, Reinisch N, Braunsteiner H: Stimulation of monocyte chemotaxis by human growth hormone and its deactivation by somatostatin. *Blood* 82:954-960, 1993
 104. Niedermühlbichler M, Wiedermann CJ: Suppression of superoxide release from human monocytes by somatostatin-related peptides. *Regul Pept* 41:39-47, 1992
 105. Grant MB, Wargovich TJ, Ellis EA, et al: Localization of insulin-like growth factor I and inhibition of coronary smooth muscle cell growth by somatostatin analogues in human coronary smooth muscle cells. A potential treatment for restenosis? *Circulation* 89:1511-1517, 1994
 106. Simons M, Edelman ER, DeKeyser JL, et al: Antisense *c-myc* oligonucleotides inhibit intimal arterial smooth muscle cell accumulation *in vivo*. *Nature* 359:67-70, 1992